Alpha-1 Antitrypsin Deficiency in Iranian Patients with Chronic Obstructive Pulmonary Disease

Bitā Geramizadeh^{1,2,*}, Zahra Jowkar¹, Leila Karami¹, Masoum Masoumpour³, Samrad Mehrabi³, Mohammad-Ali Ghayoumi³

¹Department of Pathology, Transplant Research Center, Shiraz University of Medical Sciences, Shiraz, IR Iran

²Transplant Research Center, Shiraz University of Medical Sciences, Shiraz, IR Iran

³Department of Internal Medicine, Shiraz University of Medical Sciences, Shiraz, IR Iran

*Corresponding Author: Bitā Geramizadeh, Department of Pathology, Transplant Research Center, Shiraz University of Medical Sciences, Shiraz, IR Iran. Tel: +98-7116474331, Fax: +98-7116474331, E-mail: geramib@sums.ac.ir.

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Background: Alpha-1 antitrypsin deficiency is a genetic disease which affects both lung and liver. This disease is a recognized factor for chronic obstructive pulmonary disease (COPD). However its importance as the cause of COPD in a country such as Iran is unclear.

Objectives: This study was conducted to find out the role of α -1 antitrypsin deficiency as a cause of COPD in Iranian patients.

Materials and Methods: The serum concentration of α -1 antitrypsin was determined and the genotype of α -1 antitrypsin was also evaluated by PCR-RFLP in 130 patients with COPD and 50 normal healthy blood donors.

Results: No α -1 antitrypsin deficient case was found in normal healthy people and COPD patients.

Conclusions: Our results clarify that deficiency of α -antitrypsin is not a major cause of COPD in Iranian patients.

Keywords: Alpha-1 Antitrypsin Deficiency; COPD; Iran

1. Background

Alpha-1 antitrypsin (AAT) is an acute phase reactant mostly produced by the liver and its major function is to inhibit neutrophil elastase and protect lungs against the enzyme action (1). AAT gene is located on chromosome 14q32. It is a highly polymorphic gene with over 100 alleles identified, at present (2). The SERPINA1 PI*M allele codes for the normal AAT variant, whereas PI*S and PI*Z are the most common deficiency alleles. Deficiency alleles other than PI*S and PI*Z have very rarely been reported (1). AAT deficiency is associated with severe pulmonary emphysema and chronic obstructive pulmonary disease (COPD) (3). The prevalence of AAT in COPD patients varies from one country to another (4). WHO has advised screening for AAT in all COPD patients once in their life (5). To the best of our knowledge there is no published study in the English literature from Iran about the prevalence of AAT deficiency in COPD patients.

2. Objectives

We conducted this study to find out the role of this defi-

ciency as a cause of COPD.

3. Material and Methods

During a one year period (2009 - 2010), 130 patients who referred to with the symptoms of COPD (cough, dyspnea, wheezing and shortness of breathing) and showed FEV1 < 80%, which is the diagnostic criteria for COPD, as recommended by American society of thoracic diseases were selected for the study (6). For each patient 10 cc of blood (5 cc clot and 5cc in EDTA) was collected.

DNA was isolated from the whole blood (in EDTA) using phenol-chloroform procedure and was frozen at -20°C. PCR-RFLP (Polymerase chain reaction-restriction fragment polymorphism) was performed with the below-mentioned primers. Primers for Z mutation were as below:

PFZ: 5-ATAAGGCTGTGCTGACCATCGTC-3 (T_m: 65°C)

PRZ: 5-TTGGGTGGGATTCACTTTTC-3 (T_m: 63°C)

Primers for S mutation were as below:

PRS: 5-TGAGGGGAACTACAGCACCTCG-3 (T_m: 66°C)

PRS: 5-AGGTGTGGGCAGCTTCTTGGTCA-3 (T_m: 66°C)

The PCR reaction mixture is shown in Table 1.

Implication for health policy/practice/research/medical education:

This study would help Internists and pulmonologist to understand that alpha-1 antitrypsin deficiency is not a major cause of COPD in Iranian Patients

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Table 1. The PCR Mixture

PCR	Volume, tub, μ L	Final Concentration
10 X PCR buffer	5	1X
50 mM mgcl ₂	1.5	1.5 mM
10mM dNTPS	1	200 μ M
10mM PF&PR	2.5	0.5 μ M
5 unit / μ l poly-merase	0.5	2.5 unit
Distilled water	32	-
Template DNA	5	-

The PCR temperature and cycles are shown in Table 2.

Table 2. The PCR Temperature and Cycles

Round	Profile	Denaturation	Annealing	Extension	None of Cycles
First	Temperature, °C	94	-	-	1
	Time, min	10			
Second	Temperature, °C	94	60	72	35
	Time, s	2	2	2	
Third	Temperature, °C	-	-	72	1
	Time, min	-	-	10	

Figure 1. S Mutation

Lane 1, MM Genotype: 100bp (after enzyme); Lane 2, PCR product: 121bp (before enzyme); Lane 3, 50bp ladder; Z mutation: Lane 4, MM Genotype: 157bp (after enzyme); Lane 5, PCR product: 179bp (before enzyme)

In the meantime, the level of AAT was measured in the serum by nephelometry. (Minineph TM human AAT kit, product code ZK034R).

4. Results

130 COPD patients were included in this study. The age range was 21 to 70 years (mean = 48.7 ± 8). 126 patients were male and only four females were selected. 125 patients were smokers.

59 healthy blood donors with normal respiratory func-

This schedule was the same for S and Z primers.

To test for Z and S alleles' mutation, an aliquot of the PCR product (20 μ L) was digested with 10 units (taq1) of restriction enzyme and 3 μ L of 10X buffer (on ice), then distilled water was added to reach final volume of 30 μ L. After mixing gently and centrifuging at 3000 rpm for 20 seconds, the sample was incubated at 65°C for 16 hours. Upon digestion, 10 μ L of cleaved product was submitted to electrophoresis in 3% agarose gel for one hour and stained by Ethidium Bromide.

Normal alleles will be digested by the taq1 enzyme resulting in 157bp and 22bp fragments, but the enzyme will not digest the mutant alleles. Water was used as a negative control (Figure 1).

tion were also selected for the control group with mean age of 28.4 years (51 males and 8 females). The AAT level by nephelometry was normal in all patients and controls (0.629 - 1.696 gr/L). All patients and controls showed MM genotypes and no mutant cases were detected. The PCR products were also analyzed by DNA sequencing and this analysis also confirmed the MM genotype. So in these COPD and healthy groups of people, no case of AAT mutation and deficiency was identified.

5. Discussion

AAT is a 52 KD glycoprotein mostly secreted by hepatocytes but also secreted by other cells such as lung epithelial cells and phagocytes (7). The specific target of AAT is neutrophil elastase, an enzyme that digests elastin, basement membrane and other extracellular matrix components (8). AAT deficiency causes predisposition to develop a number of diseases through life, mainly COPD and several types of liver diseases (4). PiMM is the normal and most common allele, but AAT gene is highly polymorphic with more than 100 alleles. Variants are classified according to the protease inhibitor (Pi) system (9). Most of the pathology is related to the Z allele and more than 96% of the patients worldwide have a ZZ phenotype and the remaining 4% mostly belong to the SZ and MZ variants (4).

Prevalence of AAT deficiency varies from one country to another and knowledge about the AAT deficiency status in every country is essential (10). It is estimated that

34 million people worldwide are deficient for AAT (PiZZ, PiSZ, PiSS) and 116 million are carriers (11).

AAT deficiency is mostly prevalent in European countries with high frequencies in Spain and Portugal (11). There are very few studies on AAT deficiency as a cause of COPD from Asian countries, most of which have concluded that AAT deficiency is a rare cause of COPD in these countries (1, 8, 12-14). In Hong Kong, a study on 356 COPD patients with mean age of 72.4 years old and 185 healthy unrelated controls identified 63% homozygote for M type and 4% heterozygote for M type and none were homozygous for any deficient allele (14). Based on a study on Korean patients with COPD, involving 114 patients and 196 healthy controls, same distribution of variants in both groups was identified, but S and Z alleles were not found (11). According to another study in Korea on 56 male emphysematous patients above 50 years old serum level of AAT was assayed, also phenotyping and genotyping were done; no S or Z variant was identified and M variant among COPD patients and healthy group was similar (12). In Saudi Arabia, in 158 healthy individuals, genotyping was done which showed 2.53% were heterozygous for Z mutation, 11.39% were heterozygous for S mutation and 3.8% for SZ. Homozygous for SS was present in 1.9% of individuals and no ZZ phenotype was detected (8). In China, serum analysis and isoelectric focusing electrophoresis of 748 normal individuals and 414 COPD patients did not find PiZ or PiS mutations in both groups and suggested that AAT deficiency is not a factor in the development of COPD (13). In Turkey, investigation on 123 healthy blood donors by isoelectric focusing and nephelometry determined no Z and S homozygous variant (15). In India, genotyping by PCR-RFLP technique on 200 COPD patients showed only 2 SS allele (1%) and ZZ variant (0.5%) (16). Based on these studies and current the study in Iran, it seems that AAT is not a common cause of COPD development in the Asian population. However, there is no published study in the English literature about the frequency of AAT deficiency among COPD patients from Iran. According to our study AAT deficiency is definitely not a major cause of COPD in Iran which is the same as other Asian countries.

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Authors' Contribution

None declared

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References

1. Denden S, Zorzetto M, Amri F, Knani J, Ottaviani S, Scabini R, et al. Screening for Alpha 1 antitrypsin deficiency in Tunisian subjects with obstructive lung disease: a feasibility report. *Orphanet J Rare Dis*. 2009;**4**:12.
2. Carroll TP, O'Connor CA, Floyd O, McPartlin J, Kelleher DP, O'Brien G, et al. The prevalence of alpha-1 antitrypsin deficiency in Ireland. *Respir Res*. 2011;**12**:91.
3. Spinola C, Brehm A, Spinola H. Alpha-1-antitrypsin deficiency in the Cape Verde islands (Northwest Africa): High prevalence in a sub-Saharan population. *Respir Med*. 2010;**104**(7):1069-72.
4. de Serres FJ, Blanco I, Fernandez-Bustillo E. Estimated numbers and prevalence of Pi*S and Pi*Z deficiency alleles of alpha1-antitrypsin deficiency in Asia. *Eur Respir J*. 2006;**28**(6):1091-9.
5. Wencker M, Marx A, Konietzko N, Schaefer B, Campbell EJ. Screening for α -1-Pi deficiency in patients with lung diseases. *European Respiratory Journal*. 2002;**20**(2):319-324.
6. Stoller JK, Snider GL, Brantly ML. American Thoracic Society/European Respiratory Society statement: standards for the diagnosis and management of individuals with alpha-1 antitrypsin deficiency. *Am J Respir Crit Care Med*. 2003;**168**(7):818-900.
7. Luisetti M, Seersholm N. Alpha1-antitrypsin deficiency. 1: epidemiology of alpha1-antitrypsin deficiency. *Thorax*. 2004;**59**(2):164-9.
8. Aljarallah B, Ali A, Dowaidar M, Settin A. Prevalence of alpha-1-antitrypsin gene mutations in Saudi Arabia. *Saudi J Gastroenterol*. 2011;**17**(4):256-60.
9. Blanco I, de Serres FJ, Fernandez-Bustillo E, Lara B, Miravittles M. Estimated numbers and prevalence of Pi*S and Pi*Z alleles of alpha1-antitrypsin deficiency in European countries. *Eur Respir J*. 2006;**27**(1):77-84.
10. Spinola C, Bruges-Armas J, Pereira C, Brehm A, Spinola H. Alpha-1-antitrypsin deficiency in Madeira (Portugal): the highest prevalence in the world. *Respir Med*. 2009;**103**(10):1498-502.
11. Shim YS. Epidemiological survey of chronic obstructive pulmonary disease and alpha-1 antitrypsin deficiency in Korea. *Respirology*. 2001;**6 Suppl**:S9-11.
12. Kim CH, Yim JJ, Yoo CG, Lee CT, Kim YW, Han SK, et al. Alpha-antitrypsin genotypes in Korean patients with chronic obstructive pulmonary disease. *Respirology*. 2005;**10**(2):223-8.
13. Zhu YJ. Epidemiological survey of chronic obstructive pulmonary disease and alpha-1-deficiency in China. *Respirology*. 2001;**6 Suppl**:S13-5.
14. Kwok JS, Lawton JW, Yew WW, Chau CH, Lee J, Wong PC. Protease inhibitor phenotypes and serum alpha-1-antitrypsin levels in patients with COPD: a study from Hong Kong. *Respirology*. 2004;**9**(2):265-70.
15. Simsek H, Pinar A, Altinbas A, Alp A, Balaban YH, Buyukaskik Y, et al. Cutoff level to detect heterozygous alpha 1 antitrypsin deficiency in Turkish population. *J Clin Lab Anal*. 2011;**25**(4):296-9.
16. Sobti RC, Thakur H, Gupta L, Janmeja AK, Seth A, Singh SK. Polymorphisms in the HPC/ELAC-2 and alpha 1-antitrypsin genes that correlate with human diseases in a North Indian population. *Mol Biol Rep*. 2011;**38**(5):3137-44.